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An Electron-Conducting Cross-Linked Polyaniline-Based Redox Hydrogel, Formed in One Step at pH 7.2, Wires Glucose Oxidase

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Depending on its hydration, polyaniline (PANI) conducts charge carriers by two different mechanisms. When not hydrated, it conducts through one-dimensional (1D) bands. Such conduction requires at least 1D semicrystalline PANI.^{1,2} Hydration disrupts the parallel alignment of the chains and lowers the conduction. When dissolved, PANI behaves as a polymeric redox couple,^{3,4} and if cross-linked and hydrated, it can form an electron-conducting redox hydrogel.⁵ In redox hydrogels, electrons diffuse through electron-transferring collisions between hydrated reducible and oxidizable polymer segments.⁶ Hydration lowers the local viscosity and reduces attractive Coulombic interactions, thereby increasing segmental mobility and electron diffusion. It also increases the permeability of water-soluble ions and molecules.

Here we show that an electrostatic adduct of emeraldine-polyaniline and a polymer acid, which exhibits small band gap semiconductor-kind conductivity in the solid state, forms an electron-conducting redox hydrogel when cross-linked with a water-soluble diepoxide at neutral pH and hydrated. The cross-linked polymer triples its weight upon hydration, and the absorbed water makes it permeable to glucose. When glucose oxidase (GOx) is co-cross-linked within the hydrogel, its reaction centers are electrically wired. As a result, the PANI—GOx hydrogel catalyzes the electrooxidation of glucose at +0.3 V vs Ag/AgCl and at a current density of $225 \, \mu \text{A} \cdot \text{cm}^{-2}$.

The PANI–GOx bioelectrocatalyst was prepared at neutral pH in one pot in a single step. Methods reported by other groups were more difficult to practice. For example, a recently described high-rate electrocatalytic glucose oxidation method utilized special polyaniline-containing microrods with dissolved, rather than immobilized, GOx. The preparation of the microrods required pyrene sulfonic acid functionalization of single-walled carbon nanotubes, their embedding in aniline and polystyrene sulfonic acid, electropolymerization of the aniline in porous alumina membranes coated with a conductive gold support, and dissolving the alumina membrane. Although the steady-state glucose electrooxidation current densities were not reported for these microrods, voltammetric wave heights of 500 $\mu\text{A}\cdot\text{cm}^{-2}$ at 5 mV·s $^{-1}$ scan rate were observed.

Glucose-permeable poly(ethylene glycol) diglycidyl ether (PEGDGE) cross-linked electron-conducting redox hydrogels of Os^{2+/3+} complex, comprising polymers with poly(*N*-vinylimidazole) and partially N-alkylated poly(4-vinylpyridine) backbones, have been studied extensively.⁶ PEGDGE cross-links primary, secondary, and tertiary amines, as well as heterocyclic nitrogens, including those of polyanilines.⁸ When GOx is coimmobilized in the Os^{2+/3+} complex-comprising hydrogels, it is electrically wired, and 3D glucose electrooxidation catalysts, operating at -0.1 V vs Ag/AgCl

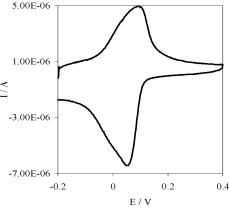


Figure 1. Cyclic voltammogram of PANI/PAAMPSA at 60 μg·cm⁻² dryweight loading; 50 mV/s scan rate; pH 7.2, 20 mM PBS.

and at >1 mA·cm⁻², are formed.^{9,10} In contrast, when GOx is physically adsorbed on,¹¹ entrapped in,¹² or electrodeposited on¹³ films of metallic or semiconducting PANI or otherwise integrated on or in PANI films,^{14–17} the glucose electrooxidation current densities are lower. Such is even the case when PANI is covalently bound to a gold substrate through a flavoenzyme thiol,¹⁸ or through nucleophilic thiol attack at ortho positions of PANI quinoimine mers.¹⁹

The emeraldine PANI was made by oxidative template polymerization of aniline on water-dissolved poly(2-acrylamido-2-methyl-1-propane sulfonic acid) (PAAMPSA) with ammonium persulfate.^{20,21} The conductivity of the PANI/PAAMPSA salt was 0.430 \pm 0.019 S/cm after the purified aqueous polymer/salt dispersion was spin- or drop-cast and dehydrated.²⁰ The molecular mass of the PAAMPSA was 724 kDa, and solid-state NMR showed that the PANI segments were 8-20 repeat units long. The shorter units were removed by added non-solvent (acetone) precipitation and by filtration and repeated washings. Elemental analysis and X-ray photoelectron spectroscopy of the purified PANI/PAAMPSA indicated an aniline/sulfonic acid molar ratio of about 1:0.9. To show that uncross-linked PANI/PAAMPSA is a typical, diffusionally mobile redox couple, glassy carbon electrodes were coated with $10 \,\mu\text{L}$ of a 0.042 wt % aqueous solution of the polymer (60 $\mu\text{g}\cdot\text{cm}^{-2}$ initial dry-weight-based loading) and allowed to dry. Figure 1 shows the initial cyclic voltammograms of the PANI/PAAMPSA-modified electrode under argon in a 20 mM, pH 7.2 phosphate buffer (PBS) at 37.5 °C. The voltammogram is characteristic of the doped PANI emeraldine redox couple, peaking at +0.1 V vs Ag/AgCl. Although the film slowly dissolved in the absence of a cross-linker, the voltammetric wave was stable at pH 7.2, and the anodic peak current increased linearly with scan rates up to 400 mV·s⁻¹, as expected for a weakly surface-bound redox couple.

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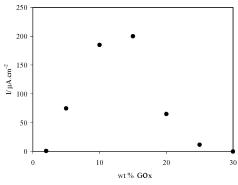


Figure 2. Dependence of the current density on the glucose oxidase weight percentage. The glucose concentration was maintained at 32 mM.

To show that a hydrogel is formed upon cross-linking the PANI/ PAAMPSA with a 400 Da PEGDGE, through reaction with PANI nitrogens,8 we measured the water uptake at 0.1 g·cm⁻² dry-weight loading of the films (cast from a 5 wt % PANI/PAAMPSA dispersion) in multiple hydration-dehydration cycles. To form the films, aqueous polymer solutions were mixed at 1:0.07 w/w PANI/ PAAMPSA solution/PEGDGE ratio, deposited on microscope slides, and cured in ambient air for 48 h. The increases-decreases in mass in cycles of (a) immersion in deionized water for 1 min, (b) tilting to drain the water, (c) contacting the surface with absorbent paper tissue (KimWipe) to remove surface water not bound in the hydrogel, (d) air-drying, and (e) repeat rehydration were measured. In three sets of measurements on different samples, the mass of the water added on swelling and lost upon drying was 2 ± 0.2 times the weight of the dry PANI/PAAMPSA/PEGDGE. To show that the hydrogel is permeable to glucose and is electrically wired to GOx, films were made by mixing droplets of known volume and concentration of aqueous solutions of PANI/PAAMPSA (in pH 7.4, 0.1 M phosphate buffer), GOx (in pH 7.2, 0.1 M phosphate buffer), and PEGDGE (in deionized water) on glassy carbon electrodes. The films were made as earlier described for an Os^{2+/3+} complex comprising redox polymer-wired GOx electrocatalyst, 10 except that the Os^{2+/3+} complex-based polymer was replaced by PANI/PAAMSA, the solution of which was now diluted 10-fold with pH 7.4, 0.1 M phosphate buffer. The films were cured for >18 h at ambient temperature.

Figure 2 shows the dependence of the glucose electrooxidation current density on the GOx weight percentage in the dry films in pH 7.2, 32 mM glucose, 20 mM phosphate buffer at 37 °C at a fixed loading of 0.8 mg·cm⁻² ($\rho = 1.39$ g·cm⁻³). In the 2–15 wt % GOx range, the current density increased with the weight percentage of GOx, reaching 200 μ A·cm⁻² at 15 wt %. At higher GOx content the current density declined. The decline is attributed to deswelling when the polyanionic GOx neutralizes the residual net positive charge of PANI/PAAMSA. To confirm that deswelling is caused by charge neutralization, GOx with a high negative surface charge was prepared by periodate oxidation of surface oligosaccharides of GOx.¹⁰ The glucose electrooxidation current density now declined when the oxidized GOx content reached 2 wt %. Figure 3 shows the dependence of the current density on glucose concentration under argon for an electrode that was poised at + 0.3 V vs Ag/AgCl in a PBS buffer. The current density was 225 μ A·cm⁻² at 40 mM glucose. The apparent Michaelis constant, K'_M, obtained from an Edie-Hofstee plot was 16.8 mM glucose.

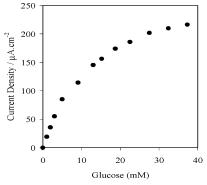


Figure 3. Dependence of the steady-state current density on glucose concentration for an electrode poised at +0.3 V vs Ag/AgCl, rotating at 500 rpm under argon.

In summary a polyaniline-based, electron-conducting, glucosepermeable redox hydrogel was formed in one step at pH 7.2 by cross-linking a polymer acid-templated PANI with water-soluble PEGDGE. Incorporation of glucose oxidase in the hydrogel by cocross-linking in the same step led to electrical wiring of the enzyme and formation of a glucose electrooxidation catalyst, allowing the electrooxidation of glucose at a current density of 225 μ A·cm⁻² at 0.3 V vs Ag/AgCl.

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References

- (1) Epstein, A. J.; MacDiarmid, A. G. J. Mol. Electron. 1988, 4, 161-165.
- (2) Lee, K.; Cho, S.; Park, S. H.; Heeger, A. J.; Lee, C.-W.; Lee, S.-H. Nature (London) 2006, 441, 65-68.
- (3) Inzelt, G. J. Electroanal. Chem. Interfacial Electrochem. 1990, 279, 169-
- (4) Horanyi, G.; Inzelt, G. Electrochim. Acta 1988, 33, 947-952.
- (5) Brahim, S.; Wilson, A. M.; Narinesingh, D.; Iwuoha, E.; Guiseppi-Elie, A. Microchim. Acta 2003, 143, 123–137.
- (6) Heller, A. Curr. Opin. Chem. Biol. 2006, 10, 664-672.
- Granot, E.; Basnar, B.; Cheglakov, Z.; Katz, E.; Willner, I. Electroanalysis
- Hua, F.; Ruckenstein, E. Macromolecules 2003, 36, 9971-9978.
- (9) Mano, N.; Mao, F.; Heller, A. J. Electroanal. Chem. 2005, 574, 347-
- (10) Mao, F.; Mano, N.; Heller, A. J. Am. Chem. Soc. 2003, 125, 4951-4957.
- (11) Chaubey, A.; Pande, K. K.; Singh, V. S.; Malhotra, B. C. Anal. Chim. Acta 2000, 407, 907–913.
- (12) Borole, D. D.; Kapadi, U. R.; Mahulikar, P. P.; Hundiwale, D. G. Polym. Adv. Technol. 2004, 15, 306-312.
- (13) Garjonyte, R.; Malinauskas, A. Biosens. Bioelectron. 2000, 15, 445-451. Parente, A. H.; Marques, E. T. A., Jr.; Azevedo, W. M.; Diniz, F. B.;
- Melo, E. H. M.; Lima Filho, J. L. Appl. Biochem. Biotechnol. 1992, 37,
- (15) Cooper, J. C.; Hall, E. A. H. Electroanalysis 1993, 5, 385-397.
- (16) Hall, E. A. H.; Skinner, N. G.; Jung, C.; Szunerits, S. *Electroanalysis* **1995**, *7*, 830–837. (17) Pan, X.; Kan, J.; Yuan, L. Sens. Actuators, B 2004, B102, 325-330.
- (18) Han, C.-C.; Heish, W. D.; Yeh, J. Y.; Hong, S.-P. Chem. Mater. 1999,
- (19) Simon, E.; Halliwell, C. M.; Toh, C. S.; Cass, A. E. G.; Bartlett, P. N. J. Electroanal. Chem. 2002, 538-539, 253-259.
- Yoo, J. E.; Cross, J. L.; Bucholz, T. L.; Lee, K. S.; Espe, M. P.; Loo, Y.-L. J. Mater. Chem. 2007, 17, 1268-1275.
- (21) Lee, K. S.; Smith, T. J.; Dickey, K. C.; Stevenson, K. J.; Loo, Y.-L. Adv. Funct. Mater. 2006, 16, 2409–2409.

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